BBA 75 284

EFFECT OF CALCIUM, POTASSIUM AND OUABAIN ON THE OXYGEN CONSUMPTION OF EXTERNAL MEDULLA SLICES FROM DOG KIDNEY

SARA WEINSCHELBAUM DE JAIRALA, LUIS GOLDMAN, ADALBERTO VIEYRA, ANTONIO P. GARCÍA AND MARTA L. RASIA

Instituto de Fisiología, Facultad de Ciencias Médicas, Universidad Nacional de Rosario, Rosario (Argentina)

(Received January 16th, 1969)

SUMMARY

Slices of external medulla from dog kidney were incubated in Krebs–Ringer solution with variable concentrations of Ca^{2+} ([Na⁺] and [K⁺] remaining constant) and K⁺ ([Na⁺] and [Ca²⁺] remaining constant). It was found that K⁺ and Ca^{2+} stimulated Q_{02} at low concentrations but inhibited it at high concentrations. The stimulation phase was studied for both cations by using three different Na⁺ concentrations. Ouabain inhibited the Q_{02} of external medulla slices of kidney and this inhibition diminished with increasing concentrations of K⁺ in the extracellular fluid. According to these results the interaction between K⁺ and ouabain seemed to be competitive.

INTRODUCTION

The oxidative exergonic reactions occurring in the external medulla are essential for the formation of the renal interstitial osmotic gradient which is primarily made up of Na⁺ accompanied by the chloride ion and urea. The present paper deals with the influence of Ca²⁺, K⁺ and ouabain on the $\rm O_2$ consumption of external medulla slices from dog kidney. This $\rm O_2$ consumption is basically associated with an active reabsorption of Na⁺ (refs. 1–3); and, for this reason, the interaction of the previously mentioned agents with Na⁺ has been investigated.

MATERIAL AND METHODS

Dogs were shot in the head, and the kidneys were removed immediately. After being placed on iced glass, the external medulla was cut with a razor blade. The slices were incubated in Krebs-Ringer solution buffered with Tris which also contained glucose and α -ketoglutarate⁴.

The total osmolarity of the incubation medium was kept constant by adding choline chloride when needed. The Q_{02} was measured in a Warburg respirometer and was expressed in μ l O_2 /mg dry tissue per h and was transformed as fractions of basal consumption (consumption in a medium without sodium).

RESULTS

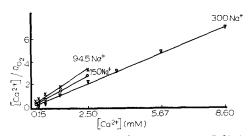
Calcium

Slices of external medulla incubated in Krebs-Ringer solution without Ca^{2+} showed a significant lowering in O_2 consumption⁵. In order to study the influence exerted by different extracellular Ca^{2+} concentrations, the Q_{O_2} was measured by incubating the slices in a Krebs-Ringer solution containing a constant Na^+ concentration (300 μ equiv/ml) and variable Ca^{2+} concentrations (from 0 to 128.4 μ equiv/ml).

When the Ca²⁺ concentration was raised from 0 to 25 μ equiv/ml, a sharp increase in O₂ consumption was obtained, while from 25 to 128.4 μ equiv/ml, the Q_{O_2} decreased with a shallow slope. The process of stimulation of oxygen consumption was also studied under a constant Na⁺ concentration of 150 μ equiv/ml and 94.5 μ equiv/ml.

Hyperbolic curves were obtained when $Q_{\mathbf{O_2}}$ was plotted against $[\mathrm{Ca^{2+}}]$ at the three Na⁺ concentrations mentioned above. They could be transformed into straight lines by plotting $[\mathrm{Ca^{2+}}]/Q_{\mathbf{O_2}}$ against $[\mathrm{Ca^{2+}}]$ (Fig. 1).

It seemed interesting to investigate whether the correlation between total O_2 consumption and Ca^{2+} concentration was also maintained between suprabasal* Q_{O_2} and $[Ca^{2+}]$. By plotting $[Ca^{2+}]$ /suprabasal Q_{O_2} against $[Ca^{2+}]$ for the three Na⁺ concentrations used, high correlations were obtained (Fig. 2). The slopes and intercepts of these regressions were linearly correlated with $I/[Na^+]$ (Figs. 8 and 9).



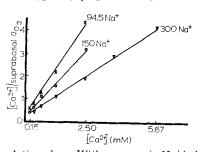


Fig. 1. Regressions of $[Ca^{2+}]/total Q_{02}$ on $[Ca^{2+}]$ in the stimulation phase. With 300 μ equiv Na⁺/ml, r = 0.96 (P < 0.001); with 150 μ equiv Na⁺/ml, r = 0.99 (P < 0.001); with 94.5 μ equiv Na⁺/ml, r = 0.99 (P < 0.001).

Fig. 2. Regression of [Ca²+]/suprabasal Q_{02} on [Ca²+] in the stimulation phase. With 300 μ equiv Na+/ml, r=0.95 (P<0.001); with 150 μ equiv Na+/ml, r=0.94 (P<0.001); with 94.5 μ equiv Na+/ml, r=0.91 (P<0.001).

When the correlation between [Ca²⁺] and Q_{02} was studied in the inhibition phase of the curve (25 μ equiv/ml<[Ca²⁺]<128.4 μ equiv/ml; [Na⁺] = 300 μ equiv/ml), a hyperbolic function was obtained.

By plotting $[Ca^{2+}] \cdot Q_{O_2}$ against $[Ca^{2+}]$, a straight line was obtained (Fig. 3, A). The correlation was also maintained when plotting $[Ca^{2+}] \cdot \text{suprabasal } Q_{O_2}$ against $[Ca^{2+}]$ (Fig. 3, B).

Potassium

It has been reported that a lack of K^+ in the extracellular fluid causes a significant lowering in Q_{02} in rat kidney cortex slices. In order to discover whether the

^{*} Suprabasal $Q_{\mathbf{O_2}}=$ total $Q_{\mathbf{O_2}}$ minus $Q_{\mathbf{O_2}}$ obtained in a medium without sodium.

same phenomenon is observable in the external medulla, suitable slices were incubated in Krebs-Ringer both with 5 μ equiv K⁺/ml and without K⁺ during 10 min at 37° (10 ml of Ringer per 100 mg wet tissue) and were washed with the corresponding solution. (These steps were designed to remove the bulk of that ion which might have diffused into the medium during the K⁺-free incubation). It was found that depletion of external K⁺ causes a definite lowering in Q_{02} (F = 19; P < 0.001).

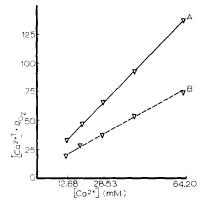
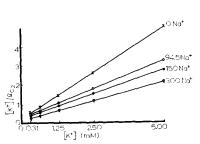


Fig. 3. Ca²⁺: inhibition phase. (A) Regression of $[Ca^{2+}]$ -total Q_{02} on $[Ca^{2+}]$; with 300 μ equiv Na⁺/ml, r = 0.98 (P < 0.001). (B) Regression of $[Ca^{2+}]$ -suprabasal Q_{02} on $[Ca^{2+}]$; with 300 equiv Na⁺/ml, r = 0.96 (P < 0.001).

The effect of variable K⁺ concentrations on Q_{O_2} was studied by working with a constant Na⁺ concentration of 300 μ equiv/ml. O₂ consumption increased as the K⁺ concentration was raised from 0 to 5 μ equiv/ml, while larger concentrations induced a decrease in Q_{O_2} . In the stimulation phase, a hyperbolic curve was obtained when correlating Q_{O_2} with [K⁺]. This was transformed into a straight line by plotting [K⁺]/ Q_{O_2} against [K⁺]. Similarly, straight lines were obtained when the medium had 94.5 μ equiv/ml and 150 μ equiv/ml Na⁺ (Fig. 4). The effect of K⁺ on Q_{O_2} in the inhibition phase was studied under 300 μ equiv/ml Na⁺; a hyperbolic function was obtained (Fig. 5).

As in the case of Ca²⁺, the existence of a correlation between [K⁺]/suprabasal



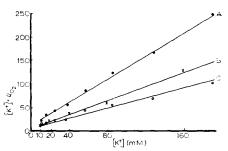


Fig. 4. Regressions of [K⁺]/total Q_{02} on [K⁺]. With 300 μ equiv Na⁺/ml, r=0.96 (P<0.001); with 150 μ equiv Na⁺/ml, r=0.98 (P<0.001); with 94.5 μ equiv Na⁺/ml, r=0.99 (P<0.001); without Na⁺, r=0.99 (P<0.001).

Fig. 5. K⁺: inhibition phase. (A) Regression of [K⁺]·total Q_{02} on [K⁺]; with 300 μ equiv Na⁺/ml, r=0.93 (P<0.001). (B) Regression of [K⁺]·basal Q_{02} on [K⁺]; without Na⁺, r=0.98 (P<0.001). (C) Regression of [K⁺]·suprabasal Q_{02} on [K⁺]; with 300 μ equiv Na⁺/ml, r=0.79 (P<0.001).

 $Q_{\mathbf{O}_2}$ and $[\mathbf{K}^+]$ was investigated. Using suprabasal $Q_{\mathbf{O}_2}$ instead of total $Q_{\mathbf{O}_2}$, the correlation obtained was insignificant (P>0.05). The basal $Q_{\mathbf{O}_2}$ was measured by the O_2 consumption in a medium without Na⁺ but rather with 5 μ equiv/ml of K⁺; therefore, K⁺ probably could also influence the $Q_{\mathbf{O}_2}$ obtained in a medium without Na⁺.

In order to investigate this point, the Q_{02} was measured while varying K^+ concentration in a medium without Na⁺. It was found that when the K^+ concentration was raised from 0 to 10 μ equiv/ml, an increase in O_2 consumption was obtained; from 10 to 160 μ equiv/ml, there was a decrease in Q_{02} . Both curves represented hyperbolic functions and were rectified by plotting $[K^+]/Q_{02}$ against $[K^+]$ for the stimulation phase (Fig. 4) (0 μ equiv/ml $<[K^+]<$ 10 μ equiv/ml) and $[K^+]\cdot Q_{02}$ against $[K^+]$ (Fig. 5) (10 μ equiv/ml $<[K^+]<$ 160 μ equiv/ml) for the inhibition phase.

The value calculated from the regression line was then considered as basal oxygen consumption for every K⁺ concentration used.

In the stimulation phase, by plotting $[K^+]$ /suprabasal Q_{O_2} against $[K^+]$ a good correlation was obtained for the three Na⁺ concentrations (Fig. 6). The slopes and intercepts of these regressions were linearly correlated with $1/[Na^+]$ Figs. 8 and 9).

In the inhibition phase, the suprabasal Q_{O_2} was calculated by deducing from the total O_2 consumption the Q_{O_2} obtained for every K^+ concentration in a medium without Na⁺. The values obtained, when multiplied by $[K^+]$ plotted against $[K^+]$, gave straight lines (Fig. 5).

Ouabain

It has been reported in different systems^{7,8} that ouabain interferes with Nartransport and lowers the $\rm O_2$ consumption.

The interaction of K^+ and ouabain in external medulla slices from dog kidney has also been investigated.

The $Q_{\mathbf{O}_2}$ of slices incubated in a medium with ouabain ($10^{-3}-4\cdot10^{-7}$ mmoles/ml) and [Na⁺] (300 μ equiv/ml), was found to be lower than in the controls ($Q_{\mathbf{O}_2}$ without ouabain = 2.43; $Q_{\mathbf{O}_2}$ with $4\cdot10^{-7}$ mM ouabain = 1.23).

The interaction between K^+ and ouabain was studied by incubating slices in a medium with 300 μ equiv/ml Na⁺ and $4\cdot 10^{-7}$ mmoles/ml ouabain. The K⁺ concentration varied from 5 to 80 μ equiv/ml. A definite rise in Q_{O_2} was observed up to 20 μ equiv/ml K⁺ followed by an inhibition of Q_{O_2} from 20 to 80 μ equiv/ml (Fig. 7).

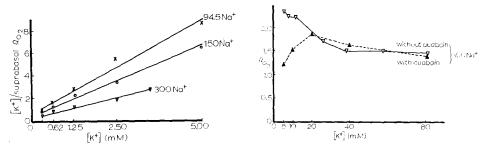


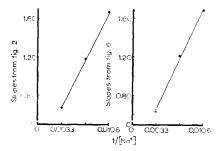
Fig. 6. Regressions of [K⁺]/suprabasal Q_{02} on [K⁺]. With 300 μ equiv Na⁺/ml, r=0.96 (P<0.001); with 150 μ equiv Na⁺/ml, r=0.96 (P<0.001); with 94.5 μ equiv Na⁺/ml, r=0.95 (P<0.001).

Fig. 7. Relationship between Q_{0_2} and $[K^+]$ in the absence and the presence of $4 \cdot to^{-7}$ mmoles/ml ouabain. $[Na^+] = 300 \ \mu equiv/ml$.

In the absence of ouabain, a lowering in Q_{02} is observed by varying the K⁺ concentration from 5 to 20 μ equiv/ml.

The stimulation phase caused by raising the K^+ concentrations with a constant concentration of ouabain may be compared with that obtained in the absence of ouabain. When $[K^+]/Q_{02}$ is plotted against $[K^+]$, both regressions have the same slope (F = 0.03; P > 0.10) but different intercepts (t = 3.66; P < 0.001).

Also the inhibitory phase in the presence of ouabain may be compared with that obtained in its absence; both lines have the same slope (F = 0.33; P > 0.10) and the same intercepts (t = 0.19; P > 0.10).



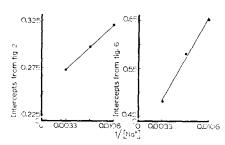


Fig. 8. Slopes from Figs. 2 and 6, plotted against 1/[Na⁺].

Fig. 9. Intercepts from Figs. 2 and 6, plotted against 1/[Na+].

DISCUSSION

The renal O_2 consumption can be divided into two parts: (1) the basal O_2 consumption which can be determined by the Q_{O_2} of kidney slices in a medium without Na⁺ (ref. 1) and in the intact mammalian kidney by the O_2 consumption in the non-filtering and hence non-reabsorbing state² and (2) the suprabasal O_2 consumption correlated with Na⁺ transportation in cortical slices¹ and also with Na⁺ reabsorption in the filtering intact kidney².

Stimulation of suprabasal Q_{O_2}

From the experimental results obtained, it could be inferred that in the absence of Ca⁺ and K⁺ in the extracellular medium the suprabasal O₂ consumption would approach zero. Whittembury⁹ and Macknight¹⁰ found that slices of kidney cortex incubated in a K⁺-free medium lost Na⁺. Kleinzeller *et al.*¹¹ incubated kidney cortex slices in the absence of Ca²⁺ and found that the slices gained Na⁺ and lost K⁺, but the Na⁺ pump was not affected by the lack of Ca²⁺. Apparently, the behavior of kidney cortex cells is different from that of external medulla slices.

In a previous work^{5,6}, a certain suprabasal O_2 consumption was obtained by using an extracellular solution without either K^+ or Ca^{2+} . It could be assumed that Ca^{2+} or K^+ , diffusing from the cells to the extracellular fluid although in a small quantity, would be sufficient to justify this small suprabasal Q_{O_2} .

The stimulating effect of Ca²⁺ on Q_{02} is in agreement with the results obtained by Dumont *et al.*¹² while measuring net Na⁺ transport in the small intestine of rats and by Bentley¹³ who reported that reduction of the Ca²⁺ concentration decreased the short-circuit current across the wall of toad bladder. And Tomlison¹⁴ also found that the short-circuit current was reduced when the Ca^{2+} concentration of the solution bathing both sides of the toad bladder was lowered. This effect of Ca^{2+} on epithelial membranes contrasts sharply, as pointed out by Blond and Whittam¹⁵, with the inhibition of Q_{02} caused by the same ion on homogenates of kidney cortex and on ATPase activity of an enzyme preparation from rabbit kidney cortex¹⁶. These differences, in effect, could be explained^{15–17} by supposing that only intracellular Ca^{2+} has an inhibitory effect. As pointed out by Opit and Charnock¹⁸, "the whole membrane structure is concerned in the transport process, although not directly involved in the chemical reaction". It seems likely, from our experimental results, that the properties of isolated enzymes could be different from the activity of the enzyme system when located in and precisely orientated at the membrane.

Inhibition of suprabasal Qo2

From the data obtained in the inhibition phase for Ca²⁺, it can be assumed that when the "stimulation" sites are filled with Ca²⁺, additional doses of this ion cause a general "stiffening" or impedance to Na⁺ transportation such as postulated by Dumont *et al.*¹² for rat small intestine, or that when the extracellular Ca²⁺ concentration is high, the intracellular Ca²⁺ concentration also increases and perhaps inactivates the Mg²⁺-dependent enzyme system.

High K⁺ concentrations inhibit ATPase activity of rabbit brain, kidney cortex¹⁹, crab nerve²⁰ and brain²¹ preparations. Apparently in all of these systems, the inhibition kinetics differ from the present experimental results. Probably the presence of intact cellular structures is likely to impose a difference on the total kinetics.

Ouabain

From the present experimental results obtained with external kidney medulla, it could be concluded that K^+ and ouabain compete for the sites in the membrane where K^+ activates the O_2 consumption or perhaps that the competitive inhibition obtained suggests that ouabain and K^+ cannot be attached to the membrane sites at the same time²¹. When all of the sites where K^+ stimulates the O_2 consumption are filled with K^+ , the inhibition by an excess of K^+ proceeds as if ouabain were absent from the system.

ACKNOWLEDGMENTS

The authors wish to express their gratitude to Dr. Sol Rabasa and Dr. Frida Bergman from the Instituto de Investigaciones Médicas, Rosario, for encouragement and helpful discussion.

Part of the work reported in these pages was supported by a grant from the Consejo para el Fomento de Investigaciones Científicas y Técnicas de la Provincia de Santa Fe.

REFERENCES

- I U. V. LASSEN AND J. H. THAYSSEN, Biochim. Biophys. Acta, 47 (1961) 616.
- 2 U. V. LASSEN, O. MUNCK AND J. H. THAYSSEN, Acta Phys. Scand., 51 (1961) 371.
- 3 K. J. Ullrich and G. Pehling, Arch. Ges. Physiol., 267 (1958) 207.

- 4 J. J. Cohen, Am. J. Physiol., 199 (1960) 560.
- 5 S. W. JAIRALA, Experientia, 21 (1965) 286.
 6 S. W. JAIRALA, Biochim. Biophys. Acta, 79 (1964) 205.
- 7 I. M. GLYNN, J. Physiol., 136 (1957) 148.
- 8 R. WHITTAM AND J. S. WILLIS, J. Physiol., 168 (1963) 158.
- 9 G. WHITTEMBURY, J. Gen. Physiol., 51 (1968) 303.
- 10 A. C. D. MACKNIGHT, Biochim. Biophys. Acta, 150 (1968) 263.
- 11 A. KLEINZELLER, A. KNOTKOVA AND S. NEDVIDKOVA, J. Gen. Physiol., 51 (1968) 326.
- 12 P. A. DUMONT, P. F. CURRAN AND A. K. SALOMON, J. Gen. Physiol., 43 (1960) 1119.
- 13 J. D. BENTLEY, J. Endocrinol., 21 (1960) 161.
- 14 J. Anderson and R. W. S. Tomlison, J. Physiol., 177 (1965) 133.
 15 D. M. Blond and R. Whittam, Biochem. J., 92 (1964) 165.
- 16 F. A. EPSTEIN AND R. WHITTAM, Biochem. J., 99 (1966) 232.
- 17 J. F. Hoffman, Circulation, 261 (1962) 120.
 18 L. J. Opit and J. S. Charnock, Nature, 208 (1965) 471.
- 19 J. C. Skou, Biochim. Biophys. Acta, 58 (1962) 314.
- 20 J. C. Skou, Biochim. Biophys. Acta, 42 (1960) 6.
- 21 K. AHMED, J. D. JUDAH AND P. G. SCHOLEFIELD, Biochim. Biophys. Acta, 120 (1966) 351.

Biochim. Biophys. Acta, 183 (1969) 137-143